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Effects of Immunomodulatory Drugs on T lymphocyte Activation and Function

Annual Report

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(AVS2149). These drugs were tested for the Pokeweed Mitogen (PWM) to lymphocyte cul						
drugs were also tested for their effects on lyn						
their ability to modulate the generation of lyn	mphokine activated	d killer cells (LA	K) which are s	timulated	d upon culturing T	
lymphocytes with Interleukin 2 (IL2). We found						
above tested functions. This inhibition was most pronounced with drug concentrations above 0.1 ug/ml. Quinolinamine at						
concentrations above 0.1 µg/ml stimulated both the PWM-stimulated IgM and IgG production, as well as the EBV-induced proliferation of lymphocytes. This drug had also a small, but significant augmenting effect on EBV-induced IgM production						
at >1 µg/ml without, however, affecting the vi	irus-induced IgG p	roduction. Fina	ally, at low cond	centration	ns (0.001-1 µg/ml)	
Quinolinamine stimulated the induction of LAK	cells while at high	er concentration	ns (2.5-20 fg/m	I) inhĺbite	ed their production.	
From the rest of the drugs poly ICLC inhibited cells, WY18251 and OK432 had a small inh	d the EBV-stimulat	ted production of	t IgM antibody	and the	generation of LAK	
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## FOREWORD

For the protection of human subjects, the investigators have adhered to policies of applicable Federal Law 45CFR46

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## BODY OF REPORT

During this third reporting year (5/15/88 to 5/14/89), the following drugs were tested for their immunomodulatory actions: Acridine trihydrochloride (CL246), OK432, WY18251, poly ICLC (AVS1761), Quinolinamine (AVS1300), and Ampligen (AVS2149). These drugs were tested for their effects on the production of antibody that is stimulated upon addition of Pokeweed Mitogen (PWM) to lymphocyte cultures or upon infection of lymphocytes with Epstein-Barr Virus (EBV). The drugs were also tested for their effects on lymphocyte proliferation stimulated by EBV. Finally, the drugs were tested for their ability to modulate the generation of lymphokine activated killer cells (LAK) which are stimulated upon culturing T lymphocytes with Interleukin 2 (IL2).

In order to study effects on mitogen-stimulated antibody production, we employed the Pokeweed Mitogen (PWM)-driven system where peripheral blood lymphocytes are stimulated in vitro and the produced antibody is measured in the tissue culture supernatant by ELISA. In particular, duplicate 12x75 mm tissue culture tubes containing 1x10<sup>6</sup> peripheral blood lymphocytes in 1 ml of RPMI 1640-10% FCS and an optimal amount of PWM (in our experiments 0.1  $\mu$ g/ml) are incubated for 9 days in a humidified atmosphere of 95% air-5% CO<sub>2</sub> at 37<sup>0</sup>C. The culture supernatants are collected, cleared of any cells by centrifugation and their IgM and IgG antibody content tested by ELISA. In the ELISA assay duplicate microtiter plate wells are pre-coated with either goat anti-human IgM or IgG (1  $\mu$ g/ml), the samples are added and then developed with a pre-determined optimal concentration of Alkaline Phosphatase (AP)-conjugated goat anti-human IgM or IgG respectively in the presence of substrate. The amount of antibody in each sample is then quantified by assessing the amount of colored end-product by optical density scanning of the plate at 405 nm.

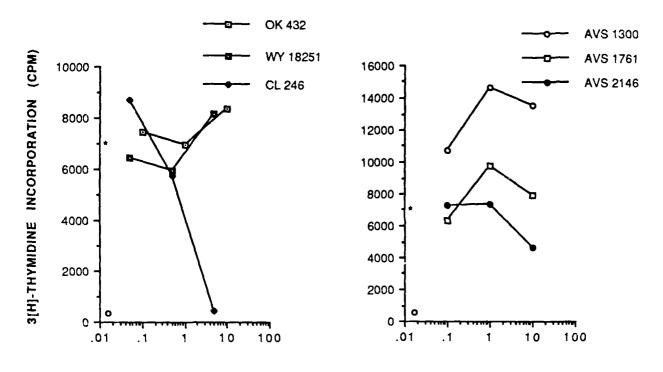
To test the effects of the drugs, peripheral blood lymphocytes were incubated with various concentrations of each drug and 0.1  $\mu g/ml$  PWM or medium as a control. The secreted IgM and IgG was then quantified by ELISA. The results shown in Figure 1 are representative of four experiments using different blood donors. It can be seen that Acridine trihydrochloride has a dose dependent suppressive effect for both antibody isotypes. In contrast, Quinolinamine had a stimulatory effect for IgM and IgG. The rest of the drugs had no significant effects on antibody production as tested in this system. None of the drugs had toxic effects in the concentration range tested.

In the above experiments, PWM induces an antibody response which is dependent on the participation of T cells thus, any effects of the drugs could be interpreted either as effects on T or B lymphocytes. In the experiments described in this section the use of EBV addresses this question. EBV is a specific B lymphotropic virus which infects B cells and causes their proliferation and production of antibody. Thus, any effects of the drugs in the EBV system must be directed specifically to the B cell itself. In this system, human peripheral blood lymphocytes are purified by removing T cells with sheep red blood cell rosetting and monocytes by adherence on plastic dishes. This gives a population of B cells which is >90% surface Ig<sup>+</sup> and contains <1% T cells. The rest of the cells in the population are residual monocytes and some natural killer cells. This degree of purity is sufficient for the studies, considering that EBV specifically infects B cells only. The B cells are placed in culture dishes and incubated in the presence of an optimal viral dose and various drug concentrations. Appropriate positive and negative controls are also included. After an appropriate incubation period cellular proliferation is assessed and the production of antibody released in the culture supernatant is quantified by ELISA.

When the effects on EBV-induced proliferation v assessed, we found that Acridine trihydrochloride was inhibitory at concentration. The production while Quinolinamine was stimulatory at concentrations >0.1  $\mu$ g/ml (Figure 2. As for the effects on the EBV-induced antibody production, Acridine trihydrochloride again displayed a suppressive effect at similar concentrations while Quinolinamine stimulated only the IgM response, but not the IgG response (Figure 3). WY18251 and OK432 had a slight inhibitory effect on the IgM production. (Figure 3). The rest of the drugs had no detectable effects at the concentrations tested. The results with Acridine trihydrochloride and Quinolinamine are very consistent with chose obtained with PWM, suggesting that the effects of these drugs are directed against B lymphocytes.

## APPENDIX



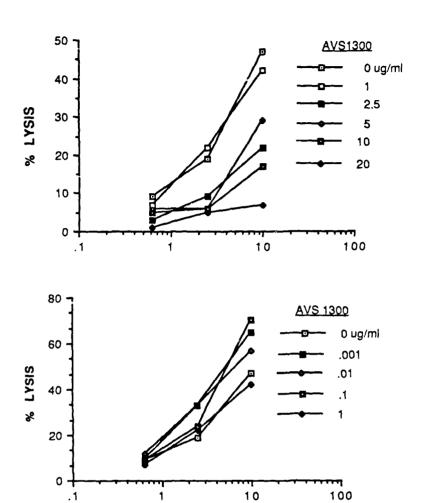


DRUG CONCENTRATION (µg/ml)

Effects of drugs on the EBV-induced proliferation of B lymphocytes: The B lymphocytes were obtained by depleting peripheral blood lymphocytes of T cells and macrophages with sheep red blood cell rosetting and plastic adherence respectively. B cells,  $2 \times 10^5$  per microtiter tray well, were cultured in total volume 0.2 mls for 8 days with or without Epstein-Barr virus (an 1:5 dilution of culture supernatant of the EBV-producer marmoset cell line B95-8) in the presence of various drug concentrations as indicated above. Cultures were incubated for 8 days and proliferation was assessed by addition of 3[H]-thymidine (1  $\mu$ Ci/well) during the last 5 hours of incubation. The "\*" indicates the positive control with EBV without any drug and the "o" indicates the negative control with only medium. The results are expressed as average CPM of triplicate cultures versus drug concentration. The standard deviation of replicate points was <10% of the average. The results are typical of four separate experiments using different donors.

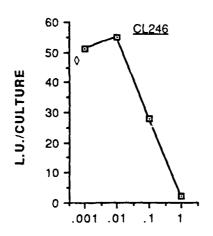
Legend to Figure 3

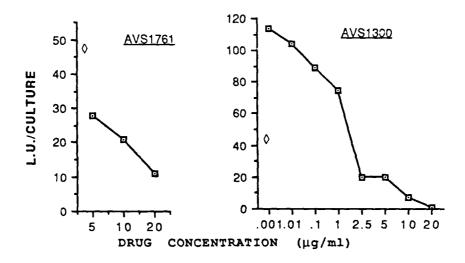
Effects of Drugs on the EBV-induced Igm and IgG production by B lymphocytes: Peripheral B lymphocytes were isolated as described in the legend of the figure above and cultured at 1x10<sup>6</sup> cells per well per ml with or without EBV (1:5 dilution of virus-containing supernatant) in the presence of various drug concentrations as indicated above. After 14 days of culture the cells were removed and the supernatants were assayed for the presence of IgM and IgG using a solid phase ELISA assay described in the text above. The "\*" indicates the positive control with EBV without any drug and the "o" indicates the negative control with only medium. The results are expressed as average 0.D. at 405 nm of duplicate determinations versus drug concentration. The standard deviation of replicate points was <5% of the average. The results are typical of four separate experiments using different donors.



See legend of Figure 4. Results are representative of three replicate studies.

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The results presented in Figures 4 and 5 have been analyzed in terms of Lytic Units (L.U.) per culture. One L.U. in this case is defined as the amount of LAK cells causing 20% lysis of the targets. The L.U. per culture were calculated from the total number of viable LAK cells recovered in each culture. The cell viability in all cultures ranged from 80-95%. The (0) represents the control culture without any drug.